

# Genetic Improvement of

# **Natural Enemies**

Senthilraja N<sup>1\*</sup>, Raghunandan BL<sup>2</sup>, Bhavik. J. Solanki<sup>1</sup> and D. B. Sisodiya<sup>1</sup>

<sup>1</sup>Department of Entomology, B. A. College of Agriculture, Anand Agricultural University, Anand, Gujarat.

> <sup>2</sup>AICRP on Biological Control of Crop Pests, Anand Agricultural University, Anand, Gujarat.

> > \*Email: rajasenthil748@gmail.com

Received: January, 2023; Revised: February, 2023 Accepted: February, 2023

# Introduction

Using natural enemies (predator, parasitoid and pathogen) to control insect pests is called biological control. A predator is an animal that preys upon other animals which are either smaller or weaker than itself. A parasitoid is a special kind of parasite which is often about the same size as its host, kills its host and requires only one host (prey) to develop into a free-living adult and the use of microorganisms like viruses, bacteria, protozoa, fungi, rickettsia and nematodes, which kill their host or debilitate future generations is called microbial control. The biological control industry in India is well organized. NBAIR act as a nodal agency for the collection, characterization, documentation, conservation, exchange, research and utilization of agriculturally important insect resources (including mites, spiders and related arthropods) and insectderived resources for sustainable agriculture. Nowadays, the application of different pesticides may depress populations of beneficial insects and also there are serious constraints to the establishment and success of biological control protocols due to environmental conditions, adaptation to crops, compatibility with pesticides, etc. Recent research has shown that, like any other system, there is scope for improvement of biocontrol agents also. Both laboratoryselected or genetically engineered natural enemies can be established in the field and enhance the efficiency of IPM programs by reduction of pesticide use.



#### Potential traits to increase the performance of natural enemies

Pesticide resistance, acceptance of food supplementation, resistance to plant defence mechanisms, starvation resistance microbiome, tolerance to relative humidity range, tolerance to extreme temperatures, resistance to desiccation, improved synchronization with the host and nondiapause are the traits to be incorporated to increase the performance of the natural enemies (Bielza et al., 2020).

# Successful improvements of natural enemies through different methods

#### 1. Artificial selection

Trichogramma chilonis strain TcT1E tolerant to endosulfan (resistance factor 9.55) and an insecticide tolerant strain PTS-8 of Chrysoperla zastrowi sillemi, having tolerance to different groups of pesticides, viz. organophosphate, organochlorine and synthetic pyrethroid was developed (NAIP, 2012). Multiple insecticide-tolerant strains of Trichogramma chilonis (MITS-TC), tolerant to endosulfan, monocrotophos and fenvalerate were developed by NBAIR. Patel and Yadav, (1995) reported the monocrotophos-resistant strain of Chrysoperla carnea. The same strain has shown cross-resistance to other insecticides such as dimethoate. acephate, phosphamidon methyl-o-demeton and (Patel and Yadav, 2000).

#### 2. Hybridization

Interspecific hybrids of *Chrysoperla* with increased fertility were developed by crossing *C. carnea* and *C. nipponensis* (Naka *et al.*, 2005). Mukuka *et al.* (2010) developed intraspecific hybrid strains of *Heterorhabditis bacteriophora* which were heat-tolerant up to 40-42 °C and desiccation tolerant. Venkatesan and Jalali (2015) developed *T. chilonis* which can tolerate



high temperatures (32-38°C) and chemical insecticides *viz.*, endosulfan, monocrotophos and fenvalerate.

#### 3. Mutagenesis

The production of the delta-endotoxin and Vip3Aa16 toxin by *B. thuringiensis* was enhanced through the mutagenesis of vegetative cells (Ghribi et al., 2004; Hmani et al., 2018). Thermotolerant mutants of Beauveria bassiana and Lecanicillium lecanii were developed using physical mutagens viz., moist heat stress (35°C) and al., UV (Avanti et 2014). Isaria fumosorosea with enhanced resistance against the broad-spectrum fungicide, benomyl was developed (Shinohara et al., 2013). The strains BB22 and BB24 with tolerance to the broad-spectrum fungicide benomyl were developed by exposing these to ion beams and gamma rays (Fitriana et al., 2015). Mutants of Heterorhabditis bacteriophora with high longevity were also developed.

4. Recombinant DNA Technology



Ecofarming e-Magazine for Agriculture and Allied Sciences http://www.rdagriculture.in e-ISSN: 2583-0791

		-		
Gene	Source	Natural Enemy	Use	Reference
Insect cuticle	Metarhizium	Metarhizium	25 per cent	Leger et al. (1996)
degrading	anisopliae	anisopliae	reduction in time	
protease (PR1)			of kill in Manduca	
			sexta	
Insect-specific	Androctonus	B. bassiana	Reduce the	Lu et al. (2008)
scorpion	australis		Median Lethal	
neurotoxin AaIT			Times against	
PR1A	M. anisopliae		Dendrolimus	
			punctatus and	
			Galleria	
			mellonella	
Carbendazim	Botrytis cinerea	L. lecanii	380-fold	Zhang <i>et al</i> .
tolerant gene			resistance to the	(2014)
(mrt)			fungicide	

#### **Other techniques**

Strains of *M. anisopliae* and *B. bassiana* with fast mycelia growth and abundant sporulation were developed by protoplast fusion (Sirisha *et al.* 2010). Development of herbicide-tolerant strains of *L. lecanii*, with tolerance to phosphinothricin herbicide with a broad spectrum of action was developed using electroporation (Timofeev *et al.*, 2019). Presnail and Hoy, (1992)

transformed the predatory mite, М. occidentalis, with a heat shock protein (HSP 70) from Drosophila and Bar gene from Streptomyces hygroscopicus (Jensen) was transferred into *M. anisopliae* var. offer resistance against acridum to herbicides like bialaphos and glufosinate ammonium (Inglis et al., 2000) using microinjection and microprojectile methods respectively.



*Trichogramma* sp Source: Arve *et al.* (2014)

# Conclusions

Natural enemies have no hazardous environmental impact, but their efficacy

*Chrysoperla* sp Source: https://entomology.ca.uky.edu/ef708

under various conditions limits their usage. The improvement of their potency can



boost their uses over the chemical mode of pest control. The impact of genetically modified organisms in farmers' fields must be assessed, considering the ecological input of different organisms. Different biological attributes such as pesticide tolerance, extending temperature and

# References

- Arve, S. S., Chavan, S. M., Toke, N. R., and Patil, D. L. (2014). Scope of genetically engineered predator and parasitoid. *International Journal of Plant Protection*. 7(1):225-231.
- Avanti, B., Balaraman, K., and Gopinath, R (2014). Development of higher temperature tolerant mutant of *Beauveria bassiana* and *Verticillum lecanii. International Journal of Life Sciences Biotechnology and Pharma Research.* 3(3):100-112.
- 3. Bielza, P., Balanza, V., Cifuentes, D. and Mendoza, J. E (2020). Challenges facing arthropod biological control: identifying traits for genetic improvement of predators in protected crops. *Pest management science*, 76(11):3517-3526.
- Fitriana, Y., Shinohara, S., Satoh, K., Narumi, I., and Saito, T (2015). Benomyl-resistant *Beauveria bassiana* (Hypocreales: Clavicipitaceae) mutants induced by ion beams. *Applied entomology and zoology*. 50(1):123-129.
- 5. Ghribi, D., Zouari, N. and Jaoua, S (2004). Improvement of bioinsecticides production through mutagenesis of Bacillus thuringiensis by UV and nitrous acid affecting metabolic pathways and/or delta-

relative humidity tolerances and altering host or habitat preferences could enhance the effectiveness of natural enemies. Hence, we should utilize these techniques to manipulate natural enemies to help our farmers.

endotoxin synthesis. *Journal of applied microbiology*. 97(2):338-346.

- Hmani, M., Boukedi, H., Ben Khedher, 6. S., Elleuch, A., Tounsi, S., & Abdelkefi-Mesrati, (2018) L Improvement of Vip3Aa16 toxin production and efficiency through nitrous acid and UV mutagenesis of Bacillus thuringiensis (Bacillales: Bacillaceae). Journal of Economic Entomology. 111(1):108-111.
- Inglis, P. W., Aragao, F. J. L., Frazao, H., Magalhaes, B. P. and Valadares-Inglis, M. C (2000). Biolistic cotransformation of *Metarhizium anisopliae* var. *acridum* strain CG423 with green fluorescent protein and resistance to glufosinate ammonium. *FEMS microbiology letters*. 191(2):249-254.
- 8. Lu, D., Pava-Ripoll, M., Li, Z. and Wang, C (2008). Insecticidal evaluation of Beauveria bassiana engineered to express a scorpion neurotoxin and a cuticle degrading protease. *Applied Microbiology and Biotechnology*. 81(3):515-522.
- Mukuka, J., Strauch, O., Hoppe, C. and Ehlers, R. U (2010). Fitness of heat and desiccation tolerant hybrid strains of *Heterorhabditis bacteriophora* (Rhabditidomorpha:



Ecofarming e-Magazine for Agriculture and Allied Sciences http://www.rdagriculture.in e-ISSN: 2583-0791

Heterorhabditidae). Journal of Pest Science. 83(3), 281-287.

- NAIP (2012). Annual report 2011-12, National Agricultural Innovation Project (NAIP)-ICAR, New Delhi. Effect of abiotic stresses on the natural enemies of crop pests: *Trichogramma, Chrysoperla, Trichoderma and Pseudomonas* and mechanism of tolerance to these stresses. 79p
- 11. Naka, H., Mitsunaga, T. and Mochizuki, A (2005). Laboratory hybridization between the introduced and the indigenous green lacewings (Neuroptera: Chrysopidae: *Chrysoperla*) in Japan. *Environmental Entomology*. 34(3):727-731.
- 12. Patel, I. S. and Yadav, D. N (1995). Susceptibility of *Amrasca biguttula biguttula* and *Chrysopa scelestes* in cotton (*Gossypium* species) to three systemic insecticides. *Indian Journal of Agricultural Sciences*.vol.65 no.4.
- Patel, I. S. and Yadav, D. N (2000). Standardization of bioassay technique for developing genetically improved monocrotophos resistant strain of green lacewing, *Chrysoperla scelestes* Banks. *Pest Management and Economic Zoology*. 8:47-51.
- Presnail, J. K., and Hoy, M. A (1992). Stable genetic transformation of a beneficial arthropod, *Metaseiulus* occidentalis (Acari: Phytoseiidae), by a microinjection technique. *Proceedings* of the National Academy of Sciences. 89(16):7732-7736.
- Shinohara, S., Fitriana, Y., Satoh, K., Narumi, I., and Saito, T (2013). Enhanced fungicide resistance in Isaria fumosorosea following ionizing

radiation-induced mutagenesis. *FEMS* microbiology letters. 349(1):54-60.

- 16. Sirisha, S., Kaur, S. G. and Palem, P. C Strain improvement (2010). of entomopathogenic fungal species Beauveria bassiana and Metarhizium by protoplast fusion. anisopliae International Journal of Applied Pharmaceutical Biology and Technology. 3(1):1135-1143.
- 17. St Leger, R. J., Joshi, L., Bidochka, M. J. and Roberts, D. W (1996). Construction of an improved mycoinsecticide overexpressing a toxic protease. *Proceedings of the National Academy of Sciences*. 93(13):6349-6354.
- Timofeev, S., Tsarev, A., Senderskiy, I., Rogozhin, E., Mitina, G., Kozlov, S. and Dolgikh, V (2019). Efficient transformation of the entomopathogenic fungus *Lecanicillium muscarium* by electroporation of germinated conidia. *Mycoscience*. 60(3):197-200.
- 19. Venkatesan T. and Jalali S. K (2015). Development, characterization and field assessment of multiple insecticides and high temperature tolerant strain of an egg parasitoid, *Trichogramma chilonis* Ishii against crop pests. Edn 1, Springer, India, pp 327-345.
- 20. Zhang, Y. J., Zhao, J. J., Xie, M. and Peng, D. L (2014). Agrobacterium tumefaciens-mediated transformation in the entomopathogenic fungus Lecanicillium lecanii and development of benzimidazole fungicide resistant strains. Journal of microbiological methods. 105: 168-173.